A stochastic model for infectious salmon anemia (ISA) in Atlantic salmon farming

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Infectious salmon anemia (ISA) is one of the main infectious diseases in Atlantic salmon farming with major economical implications. Despite the strong regulatory interventions, the ISA epidemic is not under control, worldwide. We study the data covering salmon farming in Norway from 2002 to 2005 and propose a stochastic space-time model for the transmission of the virus. We model seaway transmission between farm sites, transmission through shared management and infrastructure, biomass effects and other potential pathways within the farming industry. We find that biomass has an effect on infectiousness, the local contact network and seaway distance of 5 km represent similar risks, but a large component of risk originates from other sources, among which are possibly infected salmon smolt and boat traffic.

Keywords: biomathematics; infectious disease dynamics; spatio-temporal point processes; partial likelihood; infectious salmon anemia virus; Atlantic salmon farming

1. INTRODUCTION

World aquaculture has grown tremendously in recent decades and, along with this, a number of new diseases caused by pathogens of a wide taxonomical diversity have emerged (Murray & Peeler 2005). Infectious salmon anemia (ISA) is an infectious disease of farmed Atlantic salmon (Salmo salar). Atlantic salmon farming has been a rapidly growing industry in Norway since the beginning of the 1970s, producing 582 000 tonnes of fish with a first hand value of $1.8 billion in 2005 (Statistics-Norway 2006), making Norway a leading producer. Since the first outbreak of ISA in 1984, in Norway, the virus has caused large economic damage in the salmon farming industry in Europe and North America. Control strategies have not yet succeeded in eradicating ISA, except in Scotland (Stagg et al. 2001), therefore, it is important to identify the factors affecting the transmission of the ISA virus (ISAV). In this paper, we present a stochastic space-time model for the spread of ISA in Norway during 2002–2005.

Stochastic modelling of human and veterinary infectious diseases is a field of increasing importance. Keeling et al. (2001) modelled the foot-and-mouth epidemic in UK livestock suggesting that the most important factor in the dispersal was the distance between farms. The spread of disease in aquaculture systems is a developing field of research as witnessed by recent papers (Murray 2006; Sharkey et al. 2006; Thrush & Peeler 2006). Transmission pathways for ISAV and risk factors for ISA outbreaks are still unclear. We propose a stochastic model inspired by Keeling et al. (2001) and Diggle (2006), which captures the relative importance of the main risk factors associated with different pathways of transmission. The model investigates the relative importance of proximity to an infectious farm site by seaway distance and local contact network on the rate of transmission, as well as the importance of farm site biomass on infectiousness and susceptibility. The model also compares the rate of transmission from infectious farm sites to the rate of transmission through other potential pathways within the Norwegian salmon farming industry. The model allows precise interpretations of the parameters. We have applied the model to a unique dataset, covering the whole ISA history in Norway over the years 2002–2005 and all farm sites with a recorded biomass at any time point during this period. The dataset contains location and biomass reports from 1035 farm sites and 41 ISA infection reports. The results reveal that the majority of the total risk is likely to originate from sources other than infectious farm sites. Furthermore, seaway distance and local contact networks explain the same amount of the total risk, although having an infectious farm site at a distance from 0 to 5 km implies more risk of being infected than being in the same local contact network with an infectious farm site. The results also show

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that the biomass seem to have an effect on infectiousness. Our findings challenge the efficiency of currently implemented monitoring and regulation strategies.

2. HISTORY AND EPIDEMIOLOGY OF ISA

The first recorded outbreak of ISA occurred in 1984 in Atlantic salmon parr in a hatchery on the southwest coast of Norway (Thorud & Djupvik 1988). The disease has since been reported in Atlantic Canada (1996; Mullins et al. 1998); Scotland (1998; Rodger et al. 1998); the Faroe Islands (1999; Lynoey 2003); Chile (1999; Kibenge et al. 2001) and Maine, USA (2000; Bouchard et al. 2001). Except Chile, all these areas have experienced ISA epidemics of scales.

ISA is caused by a viral pathogen (ISAV) of the Orthomyxoviridae family (Krossøy et al. 1999). The disease is characterized by lethargy, anorexia, anemia and varying degrees of mortality (Thorud 1991). In general, clinical ISA has been associated with seawater farmed Atlantic salmon. However, Kibenge et al. (2001) isolated ISAV from clinically sick farmed Coho salmon (Oncorhynchus kisutch) from Chile. The virus has also been shown to be capable of replication in brown trout (Salmo trutta) (Nylund et al. 1995) and rainbow trout (Oncorhynchus mykiss) (Snow et al. 2001). Wild Atlantic salmon and wild brown trout have been found to be infected with ISAV (Plarre et al. 2005).

In Norway, the number of seawater Atlantic salmon farm sites with verified ISA outbreaks increased steadily from the first occurrence in 1984 to a peak in 1990, when 80 ISA cases were verified. By 1992, ISA outbreaks occurred along most of the Norwegian coast (Jarp & Karlsen 1997). The number of ISA outbreaks reduced to only one in 1992 and two in 1993. This pronounced reduction has been attributed to the implementation of various preventive measures (Thorud & Høistein 2003). Later, the number of ISA outbreaks has increased moderately: between 2001 and 2005, 31 cases were verified. By 2007, ISA outbreaks occurred along most of the Norwegian coast (Kvellestad et al. 2007). The current policy regarding minimum distance between seawater salmon farming sites is 2.5 km. Historically this has not always been followed, implying that sites may have been located closer to each other.

Thorud & Høistein (2003) describe the regulatory responses to ISA in Norway. In 1991, regulations on disinfection of effluents and secure treatment of offal from slaughtering houses and processing plants were implemented. In 1996, the authorities introduced official guidelines to deal with the outbreaks of ISA. Among others, the guidelines directed prompt destruction of fish in cages with confirmed ISA diagnosis and in other cages with clinically diseased fish, and then a full eradication of the fish population within 80 working days followed by a falling period on the site. These practices have been the regulation throughout the study period 2002–2005.

Epidemiological studies conducted in Norway identified proximity to other farm sites with ISA outbreaks as a significant risk factor. These studies indicated that seawater constituted a major pathway for the transmission of ISAV and that the virus may disperse passively with the water current through normal discharge from ISAV-infected farm sites (Vågsholm et al. 1994; Jarp & Karlsen 1997). Proximity to infected farm sites has later been confirmed as a risk factor by epidemiological studies in Canada (Gustafson et al. 2005; McClure et al. 2005). Laboratory studies have also shown that the virus can disperse through seawater (Lovdal & Enger 2002). In Norway, different farm sites with common ownership may be operated by shared staff and boats, and often from a shared wharf. Hence, there is a local contact network between farm sites described by ownership. Epidemiological studies have identified a number of farm site management and husbandry-related risk factors for ISA outbreaks, e.g. shared staff between the farm sites, divers visiting the multiple farm sites, low frequency of the removal of dead fish and low rate of treatment against sea lice (Jarp & Karlsen 1997; Hammell & Dohoo 2005; McClure et al. 2005). Low density of fish in cages has shown to reduce the susceptibility (Hammell & Dohoo 2005), but the impact of fish population size on site infectiousness and susceptibility is still unclear. Other pathways than farm-to-farm transmission have been suggested. These involve infected juvenile fish (smolt) (Nylund et al. 2007), shipment of live fish (Murray et al. 2002) and transmission facilitated by various harvesting methods (Munro et al. 2003). The importance of transmission of ISA from infectious farm sites relative to the transmission from other sources such as infected smolt and well boats is not known.

3. DATA

3.1. ISA-infected farm sites

The ISA-infected sites were identified at the National Veterinary Institute (NVI), Norway, which acts as a national and international reference laboratory for ISA diagnostics. ISA is classified as a B-category infectious disease in Norway and a list 1 disease in the EU under the Council Directive 91/67/EEC. Fish samples from all suspected outbreaks are analysed at NVI. Confirmation of a diseased population is based on a combination of clinical signs and diagnostic tests. For case sites in the present study, we used the date that fish samples were received and registered at NVI as the date of detection of ISA on the premises. There is one tie in the data, i.e. on one date there are two reported outbreaks at two different sites. The slaughtering time points were given with an approximation of three months.

3.2. Farm site biomass

The Atlantic salmon farming production cycle comprise a freshwater phase in which the fish are reared from eggs to the smolt stage, followed by a seawater phase in which smolt are moved to seawater farm sites and ongrown until slaughtering. All the outbreaks of clinical ISA have been associated with the seawater phase or the use of part seawater in freshwater hatcheries (Stagg et al. 2001). Our data include Norwegian fish farm sites (sites) in seawater with a standing stock of Atlantic salmon recorded at any point in time during the study period.
Every operator of a site holding the salmonids is required to report key production statistics on a monthly basis to the Directorate for Fisheries (DFF). We have reports on biomass on all the active sites along the Norwegian coast on a quarterly basis starting from March 2000 to December 2005, and an additional report from January 2002. Figure 1 shows the number of farms and total biomass for the period. The level change in January 2002 is artificial due to a systematic change in the license register identification system and in the procedures followed for reporting. Before 2002, the operating companies sent joint reports comprising several sites, using a different identification system for each site. When merging the old system with the new one, we could not identify the location number between 13 and 26% of the sites each quarter (overall mean 20%, decreasing in time, which explains the increasing level from March 2000 to December 2001), despite that these had a standing stock of salmon in 2000–2001. In addition, there seems to have been substantial underreporting of biomass in 2000–2001. We therefore discarded the years 2000 and 2001 in our analysis. A summary on the distribution of quarterly reports on site-specific biomass for 2002–2005 can be seen in Table 1.

3.3. Seaway distances between sites

The seaway distance (the shortest distance over seawater in kilometres) between farm sites within 100 km of each other was computed using the ArcView extension Spatial Analyst (ESRI, Redlands, CA, USA). The geographical coordinates of the sites were obtained from the aquaculture license register of DFF (www.fiskeridir.no). Distances larger than 100 km are set to infinity. A summary of the distribution of the seaway distances between the sites can be seen in Table 1.

3.4. Local contact networks

Operators of fish farming sites in Norway are predominantly companies that operate many sites. Locally, the same team of workers may operate different sites and often from a shared wharf. In addition, the sites operated by the same company may share equipment, boats, etc. This implies that sites appear in networks of contact locally. To approximate such local contact networks, a variable was compiled from the aquaculture license register identifying sites with shared ownership within municipalities. Out of a total of 431 municipalities in Norway, sites were registered in 173 municipalities. We retrieved ownership from the license register only from one point in time (September 2004) and assumed that this reflects the ownership structure throughout the study period. The variable is an approximation to the effective local contact networks, which may lead to an underestimation of this effect. A summary of the distribution of the sizes of the contact networks can be seen in Table 1. Table 2 shows the size of the local contact networks to which the ISA outbreak farms belong to.

4. A STOCHASTIC SPACE-TIME MODEL

4.1. Model specification

In this section, we describe a mathematical model for the probability that a susceptible site is infected at a time point $t$, as a function of several observable covariates: the biomass standing on the site at time $t$, the biomass of an infectious site from which the virus might potentially originate at time $t$, the distance between these two sites and the local networks they belong to. In addition, we allow for a further transmission pathway along routes which are not
Event: An event at a site means that fish at that site become infected and that the infection disperses within the fish population at that site in such a way that (at a later time) ISA becomes detectable.

Detection time delay $A$: $A$ is the detection time delay in months from an event until the infection is detected. In our study, it is fixed and deterministic, but two realistic values were compared.

ISA event time $t_i$: The time $t_i$ is the time of the $i$th event; the $i$th ISA infection is detected at time $t_i + A$.

Infectious: ISA infected site $f_j$ is infectious at time $t$ for $t_i < t < t_i + A + 1$. Susceptible: Site $f_j$ is susceptible at time $t_i$ if it is active and not infectious at time $t_i$.

The definitions of detection time delay $A$ and infectious are simplifications. $A$ comprise in fact two successive time periods; an unknown delay from event until the site becomes infectious and the period of infectiousness before the detection time. The pre-infectious period is presumably quite small compared to the latter, and is ignored here. Hence a site is infectious from the time point it becomes infectious. Furthermore, $A$ varies between cases and can only approximately be assumed as a constant. ISA has a rather long and variable incubation period. Reluctance of site managers to report ISA suspicion and different traditions at the various veterinary districts may also contribute to the variability in $A$. However, we keep $A$ fixed, but experiment with two different values. Evidence from well boat movements of salmon smolts monitored, for instance, the suppliers of smolt and the sharing of well boats.

The $m$ sites that farm salmon at least some time during the study period are denoted by $f_1, \ldots, f_m$. Let $n$ be the number of reported ISA infection events in this period. In our data $m = 1035$ and $n = 41$. The outbreak sites and their times of infection are $(f_{i_1}, t_1), \ldots, (f_{i_n}, t_i)$ such that $t_1 \leq t_2 \leq \ldots \leq t_n$, where $i \in \{1, \ldots, m\}$ and the time unit is day. The whole stock containing the ISA infected fish at site $f_j$ is slaughtered at time $s_j \geq t_i$ (after the infection is detected). The site $f_j$ has location $x_j$. The following definitions are illustrated in figure 2a.

**Active site**: A site is active at time $t_i$ if it farms salmon at time $t_i$.

**Susceptible**: Site $f_j$ is susceptible at time $t_i$ if it is active and not infectious at time $t_i$.

**Infectious**: ISA infected site $f_j$ is infectious at time $t$ for $t_i < t < t_i + A + 1$.

### Table 2. Sizes of the local contact networks within municipalities that the farm sites active in 2002–2005 belong to.

<table>
<thead>
<tr>
<th>size of local contact network</th>
<th>sites with local contact network of this size (%)</th>
<th>outbreak sites with local contact network of this size (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>no local contact network (1)</td>
<td>9</td>
<td>10</td>
</tr>
<tr>
<td>small (2–3)</td>
<td>46</td>
<td>41</td>
</tr>
<tr>
<td>medium (4–6)</td>
<td>34</td>
<td>39</td>
</tr>
<tr>
<td>large (7–9)</td>
<td>6</td>
<td>5</td>
</tr>
<tr>
<td>extra large (10–11)</td>
<td>5</td>
<td>5</td>
</tr>
</tbody>
</table>
roughly three months in a site (Stagg et al. 2001). A second line of evidence comes from the time period that salmon have been reared in seawater prior to ISA outbreaks. Epidemiological data compiled for 26 outbreaks of ISA in Norway during 2003–2005 gave a period of 13 months in seawater prior to outbreak breaks of ISA in Norway during 2003–2005 gave an average of four months between six and nine months. We varied $A$ between six and nine months for different runs of the model.

Let $\lambda_f(t)$ be the rate of transmission from site $f_j$ to site $f_i$ at time $t$. Let $n_i(t)$ be the biomass of fish in 100 ton (ton=metric ton) at site $f_i$ at time $t$, $d(x_j, x_i)$ is the seaway distance in kilometre between sites $f_j$ and $f_i$, and $k_{ji}$ is an indicator which equals 1 when sites $f_j$ and $f_i$ are in the same local contact network. Furthermore, let

$$I_j(t) = \begin{cases} 
1 & \text{if site } f_j \text{ is infectious and site } f_i \text{ is susceptible at time } t, \\
0 & \text{otherwise.}
\end{cases}$$

(4.1)

We show $I_j(t)$ for an illustrational example in figure 2b. We model the rate of transmission from site $f_j$ to site $f_i$ at time $t$ as

$$\lambda_{ij}(t) = \lambda_d(t) \exp(\alpha n_i(t) + \beta_1 n_i(t)) \cdot \left[ \exp(-\phi d(x_j, x_i)) + k_{ji} \exp(\gamma) \right] I_j(t).$$

This model assumes an exponential variation of the risk with seaway distance. Table 3 shows an overview of all the covariates with a description and their respective roles in the model. Table 4 shows an overview of all the parameters with units and explanations of their respective roles in the model.

To account for transmission pathways originating from unknown sources, we introduce a further term in the model. Let the rate of transmission $\lambda_{0i}(t)$ from sources other than farm sites to site $f_i$ at time $t$ be

$$\lambda_{0i}(t) = \lambda_d(t) \exp(\theta + \beta_2 n_i(t)) J_i(t),$$

where $J_i(t)$ is an indicator which is 1 if site $f_i$ is susceptible at time $t$. For example, infected salmon smolt and well boat effects can be captured in the parameters $\theta$ and $\beta_2$.

Following Keeling et al. (2001) and Diggle (2006), the intensity for an event at site $f_i$ at time $t$ is

$$\lambda_i(t) = \sum_{j=0}^{n} \lambda_{ij}(t).$$

(4.2)

In model (4.2), we sum up all the sources from which site $f_i$ may have been infected, given the previous history; note that the ‘0’ source is also included.

### Table 3. An overview of the covariates.

<table>
<thead>
<tr>
<th>covariate</th>
<th>unit</th>
<th>description</th>
</tr>
</thead>
<tbody>
<tr>
<td>$t_i$</td>
<td>day</td>
<td>day when site $f_i$ became infected</td>
</tr>
<tr>
<td>$n_i(t)$</td>
<td>100 ton</td>
<td>biomass of fish at site $f_i$ at time $t$</td>
</tr>
<tr>
<td>$d(x_j, x_i)$</td>
<td>km</td>
<td>seaway distance between sites $f_j$ and $f_i$</td>
</tr>
<tr>
<td>$k_{ji}$</td>
<td>no unit</td>
<td>indicator which equals 1 when sites $f_j$ and $f_i$ are in the same local contact network, otherwise 0</td>
</tr>
<tr>
<td>$I_j(t)$</td>
<td>no unit</td>
<td>indicator which equals 1 if site $f_j$ is infectious and site $f_i$ is susceptible at time $t$, otherwise 0</td>
</tr>
<tr>
<td>$J_i(t)$</td>
<td>no unit</td>
<td>indicator which equals 1 if site $f_i$ is susceptible at time $t$, otherwise 0</td>
</tr>
</tbody>
</table>

### Table 4. An overview of the parameters.

<table>
<thead>
<tr>
<th>parameter</th>
<th>unit</th>
<th>factors</th>
</tr>
</thead>
<tbody>
<tr>
<td>$\lambda_d(t)$</td>
<td>no unit</td>
<td>a time-varying risk of infection constant in space, i.e. shared by all farm sites</td>
</tr>
<tr>
<td>$\alpha$</td>
<td>(100 ton)$^{-1}$</td>
<td>the effect of increasing the biomass by 100 ton at an infectious farm site</td>
</tr>
<tr>
<td>$\beta_1$</td>
<td>(100 ton)$^{-1}$</td>
<td>the effect of increasing the biomass by 100 ton at a susceptible farm site</td>
</tr>
<tr>
<td>$\phi$</td>
<td>km$^{-1}$</td>
<td>the decay in log rate of transmission from an infectious farm site</td>
</tr>
<tr>
<td>$\gamma$</td>
<td>no unit</td>
<td>the increase in log rate of transmission from an infectious farm site to a susceptible farm site when the two farm sites are in the same local contact network, compared to when they are not</td>
</tr>
<tr>
<td>$\theta$</td>
<td>no unit</td>
<td>the log rate of transmission to a susceptible farm site from sources other than infectious farm sites in the hypothetical setting that $n_i=0$</td>
</tr>
<tr>
<td>$\beta_2$</td>
<td>(100 ton)$^{-1}$</td>
<td>the effect of increasing the biomass by 100 ton at a susceptible farm site</td>
</tr>
<tr>
<td>$A$</td>
<td>month</td>
<td>the detection time delay (not estimated)</td>
</tr>
</tbody>
</table>

in Scotland in 1998 suggest that the minimum incubation time from introduction of ISAV to the manifestation of clinical disease and mortality is roughly three months in a site (Stagg et al. 2001). A second line of evidence comes from the time period that salmon have been reared in seawater prior to ISA outbreaks. Epidemiological data compiled for 26 outbreaks of ISA in Norway during 2003–2005 gave a minimum rearing period of 6 months and a median period of 13 months in seawater prior to outbreak verification (T. M. Lyngstad 2006, National Veterinary Institute, personal communication). Given that the events occurred early in the seawater rearing phase in at least some of the outbreak sites, these observations indicate that the detection time delay $A$ amounts to at least six months. We varied $A$ between six and nine months for different runs of the model.

Let $\lambda_d(t)$ be the rate of transmission from site $f_j$ to site $f_i$ at time $t$. Let $n_i(t)$ be the biomass of fish in 100 ton (ton=metric ton) at site $f_i$ at time $t$, $d(x_j, x_i)$ is the seaway distance in kilometre between sites $f_j$ and $f_i$, and $k_{ji}$ is an indicator which equals 1 when sites $f_j$ and $f_i$ are in the same local contact network. Furthermore, let

$$I_j(t) = \begin{cases} 
1 & \text{if site } f_j \text{ is infectious and site } f_i \text{ is susceptible at time } t, \\
0 & \text{otherwise.}
\end{cases}$$

(4.1)

We show $I_j(t)$ for an illustrational example in figure 2b.
4.2. Fitting the model

Diggle (2006) proposes to estimate the parameters in (4.2) by partial likelihood. If there are no ties, the estimates of the parameters in (4.2) are obtained by maximizing the partial likelihood

$$\prod_{i=1}^{n} \prod_{k \in \mathcal{R}_i} \frac{\lambda_k(t_i)}{\sum_{l \in \mathcal{R}_i} \lambda_l(t_i)},$$

(4.3)

where $\mathcal{R}_i$ is the at-risk-set (of sites) at time $t$ such that $k \in \mathcal{R}_i$ if site $j_k$ is susceptible at time $t$. If ties occur, a modification of (4.3) is necessary. Let $t^*_1, \ldots, t^*_p$ be the distinct event times ($t^*_1 < t^*_2 \cdots < t^*_p$), $d_i$ the number of events at time $t^*_i$ and $e_i$ the set of events happening at time $t^*_i$. An approximation to the partial likelihood is

$$\prod_{i=1}^{p} \prod_{k \in e_i} \frac{\lambda_k(t^*_i)}{\sum_{l \in e_i} \lambda_l(t^*_i)}.$$  

We used the Nelder–Mead optimization algorithm (Nelder & Mead 1965) implemented in the function `optim` in the statistical software R to maximize the partial likelihood. Parameter uncertainties are based on the observed Fisher information matrix (see, for example, Pawitan 2001).

As long as the baseline hazard $\lambda_b(t)$ is non-parametric, the partial likelihood is an equivalent version of the full likelihood with respect to the estimation of the parameters (Cox 1975). The efficiency gain by using a fully parametric baseline hazard and a full likelihood to give more precise results tends to be fairly small (Kalbfleisch & Prentice 2002).

5. RESULTS AND DISCUSSION

Estimated parameters for model (4.2) are given in table 5, together with estimated 95% confidence intervals. Two separate estimations were performed, one for the detection time delay $A=6$ months and another for $A=9$ months.

At a 5% confidence level, the distance parameter $\phi$ is significantly positive for both values of $A$. Hence, the risk of a susceptible site becoming infected by an infectious site decreases with increasing seaway distance between the sites. Of particular interest is the relative importance of the distance parameter $\phi$ and the local contact network parameter, $\gamma$. The smaller the $\gamma$, the lesser the effect of local contact network. The risk implied by a distance of 0 km from an infectious farm is more than seven times the risk caused by being in the same local contact network as an infectious farm. Being in the same local contact network as a farm site with an ISA outbreak has the same effect as being at a seaway distance of $-\gamma/\phi$ km from a site with an ISA outbreak and $-\gamma/\phi=5$ for both values of detection time delay parameter $A$ (six and nine months). This means that having an infectious site at a seaway distance between 0 and 5 km implies more risk of being infected than being in the same local contact network as an infectious site. Our results are based on the exponential decay of the effect of distance on the rate of transmission $\lambda_b(t)$, which is a natural assumption (starting with Cox 1975 and including Diggle 2006). The decay is necessarily sublinear, but other functions could be assumed, possibly leading to different results.

Before the study, we suspected that susceptibility would increase with increasing biomass at the susceptible farm, i.e. $\beta_1$ and $\beta_2$ would be positive. Hammell & Dohoo (2005) looked at fish density and found that a low fish density in cages reduces the ISA susceptibility. Although biomass is not the same as density, it may capture the same risk factor. Our results for $\beta_1$ and $\beta_2$ show no clear effects of biomass on susceptibility. These parameters are not significantly different from 0 for neither values of the detection time delay parameter $A$ (six or nine months). We also suspected that infectiousness would increase with increasing biomass at the infectious site. The parameter $\alpha$ is significantly positive for both $A=6$ and 9 months indicating that of two infectious farm sites, the site with the most biomass is the most infectious. The sign of $\theta$ is not of particular interest, exp($\theta$) is the constant rate of transmission to a susceptible farm site from sources other than the infectious farm sites.

It is of interest to quantify the fraction of total risk explained by seaway distance to infectious farm sites, being in the same local contact network as infectious farm sites and sources other than infectious farm sites. For this purpose, we introduce the fraction of total risk explained by seaway distance

$$r_{\text{distance}} = \frac{\lambda_b(t) \sum_{i=1}^{n} \sum_{j \in T_i} \exp(\alpha_i t_i) \exp(\beta_1 n_1(i) + \beta_2 n_2(i)) \exp(-\phi d(i, j)) I_j(t)}{\sum_{i=1}^{n} \sum_{j \in T_i} \exp(\gamma I_j(t))},$$

the fraction of total risk explained by local contact network

$$r_{\text{network}} = \frac{\lambda_b(t) \sum_{i=1}^{n} \sum_{j \in T_i} \exp(\alpha_i t_i) \exp(\beta_1 n_1(i) + \beta_2 n_2(i)) k_j \exp(\gamma) I_j(t)}{\sum_{i=1}^{n} \sum_{j \in T_i} \exp(\gamma I_j(t))},$$

and the fraction of total risk explained by other sources

$$r_0 = \frac{\sum_{i=1}^{n} \sum_{j \in T_i} \lambda_b(t)}{\sum_{i=1}^{n} \sum_{j \in T_i} \lambda_b(t)}.$$  

Here, $m = 1035$ is the total number of sites, $T_i$ is the set of days when site $j_i$ is susceptible during the study period and $U_i$ is the set of sites which are infectious at time $t$, including the ‘0’ source.

The quantities $r_{\text{distance}}$, $r_{\text{network}}$ and $r_0$ are estimated by plugging in the parameter estimates. The results can be seen in table 6. We see that $r_0$ indicate that roughly...
70% of the total risk is explained by sources other than direct transmission from infectious farm sites. Seaway distance and local contact networks explain roughly the same fraction of the total risk.

Recall that common ownership within a municipality is used as the local contact network variable. This may not represent the local contact network accurately enough. In this sense, the local contact network effect may be underestimated. Similarly, the effect of seaway distance may be underestimated. The effective distance between two sites can be a rather complex function of seaway distance and local tidal hydrodynamics (Chang et al. 2005).

Our simplified assumption might lead to a bias towards zero estimate of the distance parameter $\phi$. The denominator in $\frac{\lambda_0(t)}{\lambda_0(t)}$, that is transmission from sources other than infected farm sites, may not represent the local contact network accurately enough. In this sense, the local contact network effect may be underestimated. Similarly, the effect of seaway distance may be underestimated. The effective distance between two sites can be a rather complex function of seaway distance and local tidal hydrodynamics (Chang et al. 2005).

We have developed a stochastic model to quantify individual risk factors in the transmission of ISA in salmon farming. Seaway distance and local contact networks are found to be factors which affect the transmission between sites, and they explain roughly the same amount of the total risk. Although these factors can be underestimated, there is strong evidence that there is a large component of the total risk caused by some other sources than transmission of the virus from infectious sites. Being in the same local contact network as an ISA event site has the same effect as being at a seaway distance of 5 km from an ISA event site. This means that there is more risk involved in having an infectious site at a seaway distance from 0 to 5 km than being in the same local contact network as an infectious site. This supports a policy which requires a minimum distance between sites, but it also means that placing two sites considerably more than 5 km apart for the purpose of minimizing the risk of ISA dispersing from one to the other is of little value if the two sites are in the same local contact network. There is evidence that biomass influences infectiousness of farms. However, the model did not detect clear effects of biomass on susceptibility. The relatively small number of ISA cases in our dataset limits the precision in our results. Furthermore, the results show that more data is needed in order to identify the transmission risk factors incorporated into $\lambda_0(t)$, that is transmission from sources other than infected farm sites. For example, data on well boat traffic and smolt suppliers would help to explain $\lambda_0(t)$. Our study highlights the need to monitor putative pathways for ISAV infection to better control the spread of ISA, such as ISAV incidence and dispersal patterns for smolt from freshwater to seawater sites and well boat traffic. Our approach can be applied further to other diseases and processes, in order to detangle basic anthropogenic factors, such as ownership networks, from natural ones. In particular, if appropriate data were available, one could study emerging diseases that impose constraints on a vigorous development of aquaculture (Murray & Peeler 2005).

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