A discrete spectral analysis for determining quasi-linear viscoelastic properties of biological materials

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The viscoelastic behaviour of a biological material is central to its functioning and is an indicator of its health. The Fung quasi-linear viscoelastic (QLV) model, a standard tool for characterizing biological materials, provides excellent fits to most stress–relaxation data by imposing a simple form upon a material’s temporal relaxation spectrum. However, model identification is challenging because the Fung QLV model’s ‘box’-shaped relaxation spectrum, predominant in biomechanics applications, can provide an excellent fit even when it is not a reasonable representation of a material’s relaxation spectrum. Here, we present a robust and simple discrete approach for identifying a material’s temporal relaxation spectrum from stress–relaxation data in an unbiased way. Our ‘discrete QLV’ (DQLV) approach identifies ranges of time constants over which the Fung QLV model’s typical box spectrum provides an accurate representation of a particular material’s relaxation spectrum, and is effective at providing a fit to this model. The DQLV spectrum also reveals when other forms or discrete time constants are more suitable than a box spectrum. After validating the approach against idealized and noisy data, we applied the methods to analyse medial collateral ligament stress–relaxation data and identify the strengths and weaknesses of an optimal Fung QLV fit.

1. Introduction

The Fung quasi-linear viscoelastic (QLV) model is a standard tool for representing the nonlinear history- and time-dependent soft-tissue viscoelasticity of biological tissues [1–4]. The QLV model provides a simple fit to stress–relaxation tests, which are preferred over standard rotational rheometry for biological tissues due to issues of gripping and anisotropy. These tests involve stretching a specimen a prescribed amount and then analysing the relaxation over time of the force needed to sustain this level of stretch. However, the confidence interval for estimation of the parameters of the ‘box’-shaped temporal relaxation function that Fung suggested in his book [1] are typically large [5], which complicates comparison of materials. Further, the usual box form of the temporal relaxation function is sufficiently restrictive that many have found the need to apply different relaxation functions [6–8] or apply different QLV representations altogether [9]. Finally, identifying when the box spectrum is too restrictive to describe a specific biological material’s time-dependent mechanical responses is a challenge [3,10–17] because, with this box spectrum, the Fung QLV model can fit relaxation data even for materials whose responses to dynamic loading it would fail to predict [18,19].

We present here a simple technique to overcome these challenges. The core of the technique is a finite series that, under special conditions, reduces to Fung’s box spectrum relaxation function.
The behaviour of a linear viscoelastic material in one dimension can be represented by the following convolution integral [20,21]:

\[
\sigma(t) = \int_{-\infty}^{t} \varphi(t, u) \frac{\partial \varepsilon}{\partial u} \, du, \quad (2.1)
\]

where \( \varphi(t, u) \) is a material modulus function, \( t \) is time, \( \sigma \) is engineering stress and \( \varepsilon \) is a linearized strain. For hereditary or non-ageing materials, this reduces to

\[
\sigma(t) = \int_{-\infty}^{t} \varphi(t - u) \frac{\partial \varepsilon}{\partial u} \, du, \quad (2.2)
\]

where the relaxation function \( \varphi(t) \) describes the mechanical character of a material, ranging from an elastic solid \((\varphi = \text{const.})\) to a Newtonian fluid \((\varphi(t) = \eta \delta(t))\), where \( \eta \) is a coefficient of viscosity and \( \delta(t) \) is Dirac’s delta function. For a material with both elastic and viscous properties, \( \varphi(t) \) must be a generalized function that defines the whole spectrum of material behaviour. Here \( \varphi(t) \) is determined in general by fitting data from an experiment such as a stress–relaxation test, in which a sample is subjected to a linear ramp at strain rate \( \dot{\varepsilon} \) over time \( 0 \leq t \leq t_p \), followed by an isometric relaxation phase over \( t_p \leq t \leq t_f \) (figure 1a,b).

2.2. Fung’s quasi-linear viscoelastic model

In a biological material, \( \varphi(t) \) is typically inadequate because the relaxation function depends upon the degree to which the material is strained. Fung’s QLV model, reviewed extensively and in more detail by others [7,8,12,14,15,18,22–27], provides a simple strain-dependent extension of (2.2) in which the temporal decay of stress is independent of strain:

\[
K(\varepsilon, t) = G(t) \sigma^{(\varepsilon)}(\varepsilon), \quad (2.3)
\]

where \( G(t) \) is a normalized function of time so that \( \lim_{t \to 0} G = G_0 \) (time-independent component of \( G \)) and \( \lim_{t \to \infty} G = 1 \) (Figure 1d); \( K(\varepsilon, t) \) represents stress and \( \sigma^{(\varepsilon)}(\varepsilon) \) represents the elastic response. Based upon the concave-up force–displacement curves common to collagenous tissues, Fung proposed using an exponential relationship that he attributes to Kenedi et al. [1,28], for the elastic stress component:

\[
\sigma^{(\varepsilon)}(\varepsilon) = A(\varepsilon^{\beta \varepsilon} - 1). \quad (2.4)
\]
The stress responses to a small strain increment, applied at time \( t \), \( G(t - u) (\partial \sigma^0(u)/\partial u) \) be summed using the Boltzmann superposition principle, so that the stress for \( t \geq 0 \) can be written as
\[
\sigma(t, u) = \int_0^t G(t - u) \frac{\partial \sigma^0(u)}{\partial u} du.
\]
where \( G(t) \) is a monotonically decreasing function \([21]\); as in the fading memory model, the recent strain history determines material response. Fung termed \( G(t) \) the ‘reduced relaxation function’ and suggested the form
\[
G(t) = \frac{1 + \int_0^\infty S(\tau)e^{-\tau/t}d\tau}{1 + \int_0^\infty S(\tau)d\tau},
\]
where \( S(\tau) \) is the following function, first proposed independently by others including Neubert \([29]\):
\[
S(\tau) = \begin{cases} C_1, & \tau_1 < \tau < \tau_2 \\ 0, & \text{otherwise} \end{cases}
\]
in which \( C, \tau_1 \) and \( \tau_2 \) are material constants to be determined experimentally (figure 1c). Substituting, one arrives at the form that is ubiquitous in the biomechanics literature:
\[
G(t) = \frac{1 + \int_0^\infty (C_1/\tau_1) e^{-\tau/t}d\tau}{1 + \int_0^\infty (C_1/\tau_1)d\tau} = \frac{1 + C(E_1(t/\tau_2) - E_1(t/\tau_1))}{1 + C \ln(\tau_2/\tau_1)},
\]
where \( E_1(t) = \int_0^\infty e^{-t/\tau}d\tau \) is the exponential integral function. This version of the Fung QLV model is predicated upon the requirements of equations (2.3), (2.4) and (2.7). Although the typical implementation focuses on small strain applications, straightforward extensions of the Fung QLV model to finite strain exist \([30–32]\). However, the suitability of the assumption of equation (2.3) must be checked carefully for any material to which these extensions are applied.

In §3, we present a simple tool to identify whether the relaxation response \( S(\tau) \) follows the box spectrum of equation (2.7), and whether this response is independent of strain (cf. equation (2.3)). We further propose a simple technique for modelling a material if these requirements prove unsuitable.

### 2.3. Schematic of the Fung quasi-linear viscoelastic model

Two graphical representations of the Fung QLV model should be mentioned. These provide a foundation for interpreting the fitting and model identification tools presented in §3.

#### 2.3.1. Standard linear solid models in series

The Fung QLV model can be represented as an infinite number of standard linear solid elements in series, modified so that linear springs are replaced by nonlinear springs (figure 2a). \( S(\tau) \) for each of those elements varies between two constants: \( S(\tau_2) < C/\tau < S(\tau_1) \). Noting that the stress in each element is equal, a relationship can be written between the first and the

\[
\sigma(t) = \begin{cases} A(e^{Bu} - 1) + Ab \int_0^t C(E_1((t - u)/\tau_2) - E_1((t - u)/\tau_1)) e^{Bu} du \\ 1 + C \ln(\tau_2/\tau_1) & (0 \leq t \leq t_f) \\ A(e^{Bu} - 1) + Ab \int_0^t C(E_1((t - u)/\tau_2) - E_1((t - u)/\tau_1)) e^{Bu} du \\ 1 + C \ln(\tau_2/\tau_1) & (t_f \leq t \leq t_f) \end{cases}
\]
or
\[
\sigma(t) = \begin{cases} 
\sigma_0(e) + \sigma_d(t) & (0 \leq t \leq t_0), \\
\sigma_0(e_p) + \sigma_d(t) & (t_0 \leq t \leq t_f).
\end{cases}
\] (2.12)

Schematically, the first term on the right-hand side of these equations represents the stress in a nonlinear time-independent (strain-dependent) spring, and the second term represents a nonlinear spring, with linear viscous damping, which is time-dependent (figure 2b). After sufficient relaxation, the second term on the right-hand side of equation (2.11) or equation (2.12) approaches 0, and \(\sigma(t)\) approaches a steady state:
\[
\sigma(\infty) = \sigma_0(e_p) = \frac{A(e^{\mu_0} - 1)}{1 + C \ln(\tau_2/\tau_1)},
\] (2.13)
where \(e_p = \dot{\varepsilon}_p\). Thus, a way to ensure that the five QLV parameters are estimated correctly from a dataset is to compare the stress asymptote in the data to \(A(e^{\mu_0} - 1)/(1 + C \ln(\tau_2/\tau_1))\) from equation (2.13) (figure 1b), assuming the isometric portion of the experiment was sufficiently long to provide an accurate estimate of \(\sigma(\infty)\).

3. Methods

3.1. Continuous quasi-linear viscoelastic spectrum
To analyse a material’s relaxation spectrum without any specific pre-assumption for \(S(\tau)\), suppose \(S(\tau)\) is
\[
S(\tau) = \begin{cases} 
h(\tau) \tau \quad & \tau_{\text{min}} < \tau < \tau_{\text{max}} \\
0 \quad & \text{otherwise},
\end{cases}
\] (3.1)
where, for \(h(\tau) = C\), equation (3.1) reduces to the Fung QLV model (cf. equation (2.7)). Plugging equation (3.1) into equation (2.6), the relaxation function can be written as
\[
G(t) = 1 + \int_0^t S(\tau)e^{-t/\tau}d\tau - 1 + \frac{\int_0^t h(\tau)e^{-(t-\tau)/\tau}d\tau}{1 + \int_0^t h(\tau)/\tau d\tau}.
\] (3.2)

Then, for a fully relaxed specimen stretched at a constant strain rate \(\dot{\varepsilon}\) over the time interval \(0 \rightarrow t_p\) and held isometrically until time \(t_p\), the stress history can be written as
\[
\sigma(t) = \begin{cases} 
\frac{A(e^{\mu_0} - 1)}{1 + \int_0^{\tau_{\text{max}}} h(\tau)/\tau d\tau} \int_0^{\tau_{\text{max}}} h(\tau)e^{-(t-\tau)/\tau}d\tau e^{\mu_0} + AB\dot{\varepsilon} \int_0^{\tau_{\text{max}}} h(\tau)/\tau d\tau & (0 \leq t \leq t_0), \\
\frac{1}{1 + \int_0^{\tau_{\text{max}}} h(\tau)/\tau d\tau} \int_0^{\tau_{\text{max}}} h(\tau)e^{-(t-\tau)/\tau}d\tau e^{\mu_0} + AB\dot{\varepsilon} \int_0^{\tau_{\text{max}}} h(\tau)/\tau d\tau & (t_0 \leq t \leq t_f).
\end{cases}
\] (3.3)

In the above equations, \(A, B, h(\tau), \tau_{\text{min}}\) and \(\tau_{\text{max}}\) are unknowns that should be estimated by fitting \(\sigma(t)\) to experimental stress–relaxation data. This, unfortunately, is an ill-posed problem. However, a broad range of techniques exists to find a solution [21,34–36] that is suitable and repeatable, if not unique.

3.2. Discrete quasi-linear viscoelastic spectrum
A discretization technique simplifies this ill-posed problem. Although several approaches exist for discretizing viscoelastic responses, including the Prony series approach, our specific objective is to arrive at an approach that yields a graphical representation to easily identify the Fung box spectrum over a particular range of loading conditions.

The approach we take begins with a discrete form of the integral \(\int_{\tau_{\text{min}}}^{\tau_{\text{max}}} h(\tau)/\tau d\tau\), with the interval \((\lambda_1, \lambda_{n+1})\) divided into \(n\) equivalent logarithmic subintervals
\[
\int_{\lambda_1}^{\lambda_{n+1}} h(\tau)/\tau d\tau = \sum_{i=1}^{n} \int_{\lambda_i}^{\lambda_{i+1}} h(\tau)d(\ln(\tau)).
\] (3.4)

For the case of \(\lambda_i\) distributed equidistantly over the range \(\lambda_1 \leq \lambda_i \leq \lambda_n\), one can further simplify to write
\[
\int_{\lambda_1}^{\lambda_{n+1}} h(\tau)/\tau d\tau = T \sum_{i=1}^{n} H_i,
\] (3.5)
where the constant \(T = \ln(\lambda_{n+1}/\lambda_1) = \ln(\lambda_2/\lambda_1)\), and \(H_i\) is the height of the \(i\)th rectangle and \(\tau_i\) is the corresponding time constant (figure 3). The spectrum \(H(\tau)\) represents a tool for model identification, which we term a ‘DQLV’ spectrum, which simplifies to the continuous Fung box spectrum when appropriate. Throughout, we use the superscript DQLV to distinguish parameters arising from a DQLV fitting from those arising from a box spectrum fitting. The discrete form of equation (3.2) is
\[
G^{DQLV}(t) = \frac{1 + T \sum_{i=1}^{n} H_i e^{-t/\tau_i}}{1 + T \sum_{i=1}^{n} H_i},
\] (3.6)

and that of equation (3.1) is
\[
S^{DQLV}(\tau_i) = \begin{cases} 
\frac{H_i}{\tau_i} \quad & \tau_i \leq \tau \leq \tau_{\text{max}} \\
0 \quad & \text{otherwise}
\end{cases}
\] (3.7)
where \(H_i\) are parameters to be fit and \(\tau_i\) and \(\tau_{\text{max}}\) as described below, are chosen to encompass a range broader than that needed to describe a material. In contrast to the Neubert [29] and Fung box spectrum models, the values of \(H_i\) need not be identical. Schematically, the DQLV model is analogous to the box spectrum model, except with a finite number of elements, \(H_i\) that represent the relaxation spectrum (figure 4). For materials that are not well fit by the box spectrum model, the insertion of equation (3.7) into equation (2.6) provides for a simple extension of the QLV model.
We note that, although the DQLV spectrum reduces to a box spectrum, it is different in that the values of \( H_n \) need not be constant. Further, although this physical meaning of \( \tau_i \) and \( \tau_j \) is analogous to these constants within a Prony series, we note that the DQLV spectrum reduces to a box spectrum when a box spectrum is indeed the correct representation of a material’s relaxation response. The model does not reduce to the spectral representation that would be obtained using the generalized Maxwell model [37].

### 3.3. Numerical fitting algorithms

As shown in equation (2.10), we can split \( G^{(DQLV)}(t) \) into time-independent and -dependent parts:

\[
G^{(DQLV)}(t) = G_{0}^{(DQLV)} + G_{d}^{(DQLV)}(t) = \frac{1}{1 + T \sum_{i=1}^{n} H_i} + \frac{T \sum_{i=1}^{n} H_i e^{-t/\tau_i}}{1 + T \sum_{i=1}^{n} H_i} \tag{3.8}
\]

Because of the nature of ill-posed problems, we expect predictions to show some deviation from a Fung box spectrum even for artificial data generated from the Fung model [36,38–40]. Thus, we used a simple regularizing criterion, which acted as a penalty against unwanted states and ensured that the fitting algorithm did not become trapped in local minima. In this approach, \( H_0 \) were smoothed by a regularization function and were identified by minimizing

\[
\sum_{j=1}^{f} (\sigma^{(exp)}_{j} - \sigma^{(DQLV)}_{j})^2 + \alpha \sum_{i=1}^{k} (H_{i-1} + 2H_i - H_{i+1})^2 \tag{3.9}
\]

where \( f \) is the number of data points, \( \sigma^{(exp)}_{j} \) are known stress data or a calculable relationship and \( \alpha \) is a regulating factor. Parameters were determined using a non-negative least-squares regression [41]. For any \( \alpha \) of the order of 1, the parameter estimates were insensitive to the specific choice of \( \alpha = 1 \) was used in the model validation studies below. \( \sigma^{(DQLV)}_{j} \) are DQLV estimates of \( \sigma^{(exp)}_{j} \): 

\[
\sigma^{(DQLV)}_{j} = \begin{cases} 
\frac{A(e^{\beta_i} - 1)}{1 + T \sum_{i=1}^{n} H_i} + \sum_{i=1}^{n} H_i e^{-t/\tau_i} e^{-\beta_i \tau_i} du & (j = 1, 2, \ldots, p), \\
\frac{A(e^{\beta_i} - 1)}{1 + T \sum_{i=1}^{n} H_i} + \sum_{i=1}^{n} H_i e^{-t/\tau_i} e^{-\beta_i \tau_i} du & (j = p, p+1, \ldots, f),
\end{cases}
\tag{3.10}
\]

where \( p \) and \( f \) are the number of data points in ramp and relaxation intervals, respectively.

### 3.4. Validation of software to estimate discrete quasi-linear viscoelastic spectra

A code to obtain the DQLV spectrum of a material from stress versus time data in a relaxation test was generated in the Matlab environment. The code is available from the authors.

Because the DQLV model represents an approximation of the real spectrum of a material [36], it was important to validate the model and check the reproducibility of the approximation. Validations were performed by using the DQLV model to fit stress–relaxation data generated using either the Fung QLV model with a box spectrum or using the generalized Maxwell model, the latter having three time constants (including the infinite time constant; e.g. [42,43]).

The stability of the model with respect to noise was then studied. Random noise was added to the simulated Fung QLV stress–relaxation data to evaluate how noise affects DQLV analysis results. We superimposed upon the data noise chosen from a uniform distribution with amplitudes of 5, 10, 15, 20 or 25% of the steady-state stress. Fifty noisy datasets were generated (10 sets for each noise percentage). Relaxation spectra, \( H(\tau) \), for these new datasets were estimated using the DQLV approach. The sensitivity to noise level was then quantified by the mean square error (MSE) of the predicted stress–relaxation compared to the underlying input stress–relaxation data.

### 3.5. Characterization of medial collateral ligament relaxation

As an example of DQLV characterization of an orthopedic tissue, we studied either the left or the right MCL from \( N = 6 \) skeletally mature female New Zealand white rabbits. Prior to dissection, knees were wrapped in saline-soaked gauze and then sealed in plastic bags and fresh frozen at \(-20^\circ\text{C}\) [44]. On the day of testing, knees were thawed to room temperature and MCLs were dissected and cut free at the insertion sites [45,46]. The geometry was standardized by cutting the ligaments into dog bone shapes with a length-to-width ratio of \( 6.8 \pm 0.8 \) (width \( 1.6 \pm 0.2 \) mm). The tissue samples were fixed in custom-made soft-tissue clamps, and the cross-sectional area was determined with a laser micrometer system (\( 1.0 \pm 0.3 \) mm²) [44,47]. Measurements were taken in three locations along the length of the tissue sample and averaged for stress calculations. The tissue sample—clamp construct was then mounted to a tensile testing machine (Enduratec Elf 3200, Bose Corporation, Framingham, MA, USA). Reflective markers were placed on the tissue sample to track strain using an optical system (VP110, Motion Analysis, Santa Rosa, CA, USA). A saline drip was used to hydrate the tissue sample and maintain temperature at a constant \( 37^\circ\text{C} \). The tissue sample was aligned along the loading axis using an \( x-y \) table and then was left unloaded for 30 min to equilibrate. Specimens were elongated to strain levels of 1.25, 2.5 and 5% and held isometrically at each level for 35 min to reach equilibrium [48]. Force data were acquired at 3 Hz throughout the ramp and 35 min relaxation intervals. The ramp time for all three samples was approximately \( 9.2 \) s; thus, the specimens’ strain rates were approximately 0.135% s\(^{-1}\), approximately 0.271% s\(^{-1}\) and approximately 0.543% s\(^{-1}\), respectively. The experimental data were fit using both the DQLV and Fung box spectrum models [5].

### 4. Results and discussion

#### 4.1. Fitting of simulated data

In the first validation study, stress–relaxation data were calculated for a Fung QLV material whose relaxation response followed a box spectrum, and the DQLV model was applied to estimate the parameters used to generate these data. The
parameters chosen were those reported by Abramowitch et al. [5] for a goat MCL that was strained to 2.76% over 18.4 s and then held isometrically for 3600 s (figure 5a,b and table 1). The DQLV model was applied using equation (3.10), which constrains the DQLV model to fit stress–relaxation data. A spectrum that approximated a box spectrum was predicted, indicating that the material would be well modelled by a box spectrum representation. Several sets of time constants and different regularization parameters \( \alpha \) were evaluated to ensure that the DQLV spectrum was repeatable. The predicted spectra were insensitive to the choice of \( \alpha \) for \( \alpha \) of the order of 1, and \( \alpha = 1 \) was adopted for all subsequent analyses. The spectrum shown in figure 5c had \( n = 100 \) time constants. Increasing \( n \) provided a better quantitative fit to the stress–relaxation data, but did not change the qualitative nature of the predicted DQLV spectrum.

An interesting feature of the DQLV analysis is that the predicted DQLV spectrum, despite its deviation from a box shape, yielded error of less than 0.01% of the peak stress when predicting stress–relaxation data generated using a flat box spectrum. Indeed, the Fung box spectrum and DQLV fits were both indistinguishable from the input relaxation data (figure 5c). The logical course of action for a DQLV spectrum such as this would thus be to adopt the simpler box spectrum fitting for subsequent analysis of this material.

We next studied whether the DQLV model could identify when stress–relaxation data should not be represented by a Fung box spectrum. The input data in this case were generated using a generalized Maxwell model [33] including two Maxwell elements in addition to a linear spring, all acting in parallel. The data chosen were those reported by Shen et al. [49] for fitting the response of a sea cucumber (Cucumaria frondosa) collagen fibril following a 20 s ramp to a strain of 20% and a subsequent 520 s isometric hold (figure 6a,b and table 2). The DQLV fitting recovered a Maxwell-type spectrum with two distinct peaks at the two non-infinite time constants used to generate the input data (figure 6c).

However, the box spectrum fitting (figure 6c), also yielded an excellent fit to the stress–relaxation data. The fitting error, as observed in the plot of the residuals (inset of figure 6b), was only slightly higher than that of the DQLV fitting. This very small difference in residual error highlights the peril of fitting a Fung QLV box spectrum in the absence of a spectral evaluation such as that which the DQLV model provides: although the box spectrum model captured the relaxation data very well, this prediction captures the frequency dependence of the material response very poorly at the extremes of the range of time constants [18,19]. As we emphasize in the sequel to this article, the errors become substantial when attempting to predict material response under dynamic loading.

Finally, the fittings were remarkably robust against Gaussian random noise for both the DQLV (figure 7a) and box spectrum (figure 7b) models. MSEs increased with noise, but were less than 2 MPa for 25% noise in both models, compared to a peak stress of about 13 MPa.
4.2. Discrete quasi-linear viscoelastic fitting of stress–relaxation data of rabbit medial collateral ligament

The stress–relaxation data for rabbit MCLs at strain levels of 1.25, 2.5 and 5% all showed a characteristic rise during stretching, then gradually decreased to plateau at about 2000 s (figure 8\(\text{a, c, e}\)). Both the DQLV and box spectrum models fit the experimental data with acceptable error, but the DQLV had higher precision (figure 8\(\text{b, d, f}\)). By comparing the DQLV spectra (figure 9\(\text{a}\)) to the Fung box spectra (figure 9\(\text{b}\)) of the three stress–relaxation tests, it is clear that the box spectra weakly estimated the lower boundary of dominant time constants of the system. The DQLV model estimation, on the other hand, was more consistent.
Moreover, the DQLV model illuminated a structurally fast time constant at about 10 s and a slow time constant at about 1000 s that were not detectable by the Fung box spectra. These observations are consistent with dynamic testing reported for other tissues [18,19,50].

The DQLV spectrum showed reasonable repeatability for the three strain levels, suggesting that the Fung QLV model’s criterion of a strain-independent reduced relaxation function would be met reasonably well for the rabbit MCL specimens tested. However, the other condition, that of a flat, box-like spectrum was not met: the continuous spectrum of equation (3.1) must have constant dimensionless height $h(t) = C$ over some range of $t_1 - t_2$, but the values of $t_1, t_2$ and $C$ varied substantially for the best fits to the three tests. Errors associated with applying the box spectrum model become evident at the lower boundary of the time constants (figure 9b).

4.3. Choosing among models

For a tissue such as the rabbit MCL studied above, a more detailed description of the spectrum would be required, especially under dynamic loading. Two logical choices are a normalized, generalized form of the Maxwell model and the DQLV model. Both are related in that they involve a Prony series, and both have strengths and limitations.

The generalized Maxwell model is an excellent tool for fitting most experimental stress–relaxation data, and can usually do so with only two or three exponential terms [37]. This is a strength because of its simplicity, but is also a limitation.
because the limited number of time constants may be inadequate to reveal either the range of relaxation mechanisms or the subtle differences between materials.

The DQLV model can represent the reduced relaxation spectrum of a material such as the rabbit MCL and is convenient for several reasons. First, estimating a DQLV spectrum is a logical first step in choosing a material model for a biological material; by using a regularizing function, the approach identifies ranges of time constants over which Fung’s continuous box relaxation spectrum provides a suitable approximation and is effective at fitting this box relaxation spectrum. Second, a DQLV spectrum identifies when discrete time constants are more effective than a box relaxation spectrum for representing damping responses and provides a reasonable material model with no further fitting. Third, the DQLV spectra from multiple strain states reveal the assumption of strain-independent relaxation (cf. equation (2.3)) is supported; for example, the MCL data (figure 9a) showed DQLV spectra that are very similar for three different strain levels, indicating that the DQLV model would be a reasonable simplification. Finally, the parameters \( \Omega_i \) are insensitive to the number of time constants chosen. Increasing the number of time constants will improve the precision of the discretization of a continuous relaxation spectrum, but our experience is that the nature of this spectrum eventually converges, becoming insensitive to further increases in the number of time constants. Application of this approach is a simple and effective way to identify material relaxation spectra in an unbiased manner from stress–relaxation data.

5. Conclusion

Application of the DQLV model is a simple and effective way to identify material relaxation spectra in an unbiased manner from stress–relaxation data. The approach identifies ranges of time constants over which Fung’s continuous box relaxation spectrum provides a suitable representation of material behaviour and is effective at fitting this box relaxation spectrum. It also identifies when discrete time constants are more appropriate than a box relaxation spectrum for representing damping responses. Although the Fung QLV model with a box spectrum can fit most stress–relaxation data, including data generated using a relaxation spectrum that differs substantially from a box spectrum, errors associated with applying the box spectrum become evident at the lower boundary of the time constants. The improvement in fit to relaxation data using the DQLV model can be substantial, especially when considering behaviour over narrow ranges of material time constants. The DQLV model was able to identify correctly spectra at particular strain levels from simple stress–relaxation tests.

**Ethics.** The study was done in compliance with the National Institutes of Health guidelines for animal care and the Institutional Animal Care and Use Committee (IACUC) at the University of Pittsburgh.

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